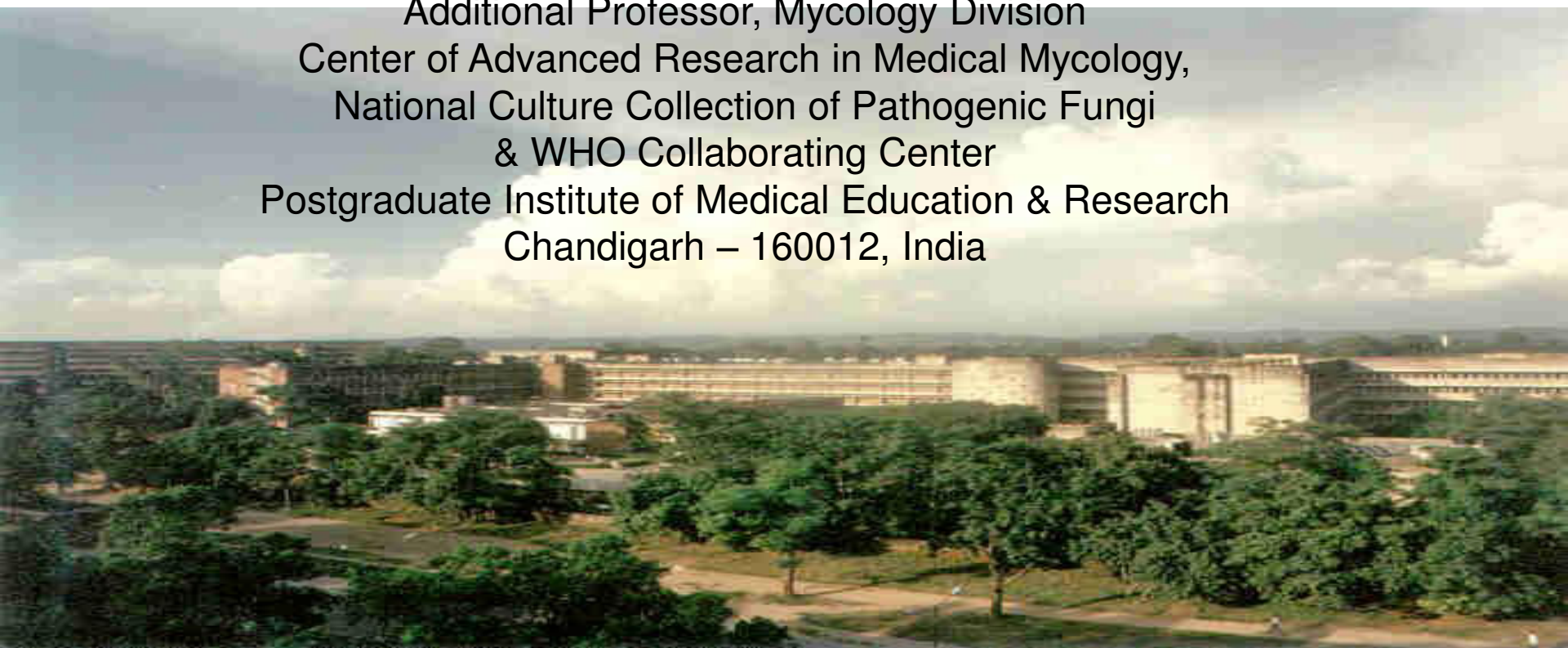


# Antifungal resistance mechanisms in pathogenic fungi

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# Antifungal drugs

## -by structure

- **POLYENES**

Amphotericin B, nystatin

- **AZOLES**

Imidazoles: Ketoconazole..

Triazoles: Fluconazole,  
itraconazole, voriconazole,  
posaconazole, ravuconazole

- **ALLYLAMINES**

Terbinafine, butenafine

- **MORPHOLINE**

Amorolfine

- **FLUORINATED PYRIMIDINE**

Flucytosine

- **ECHINOCANDINS**

Caspofungin, anidulafungin,  
micafungin

- **PEPTIDE-NUCLEOSIDE**

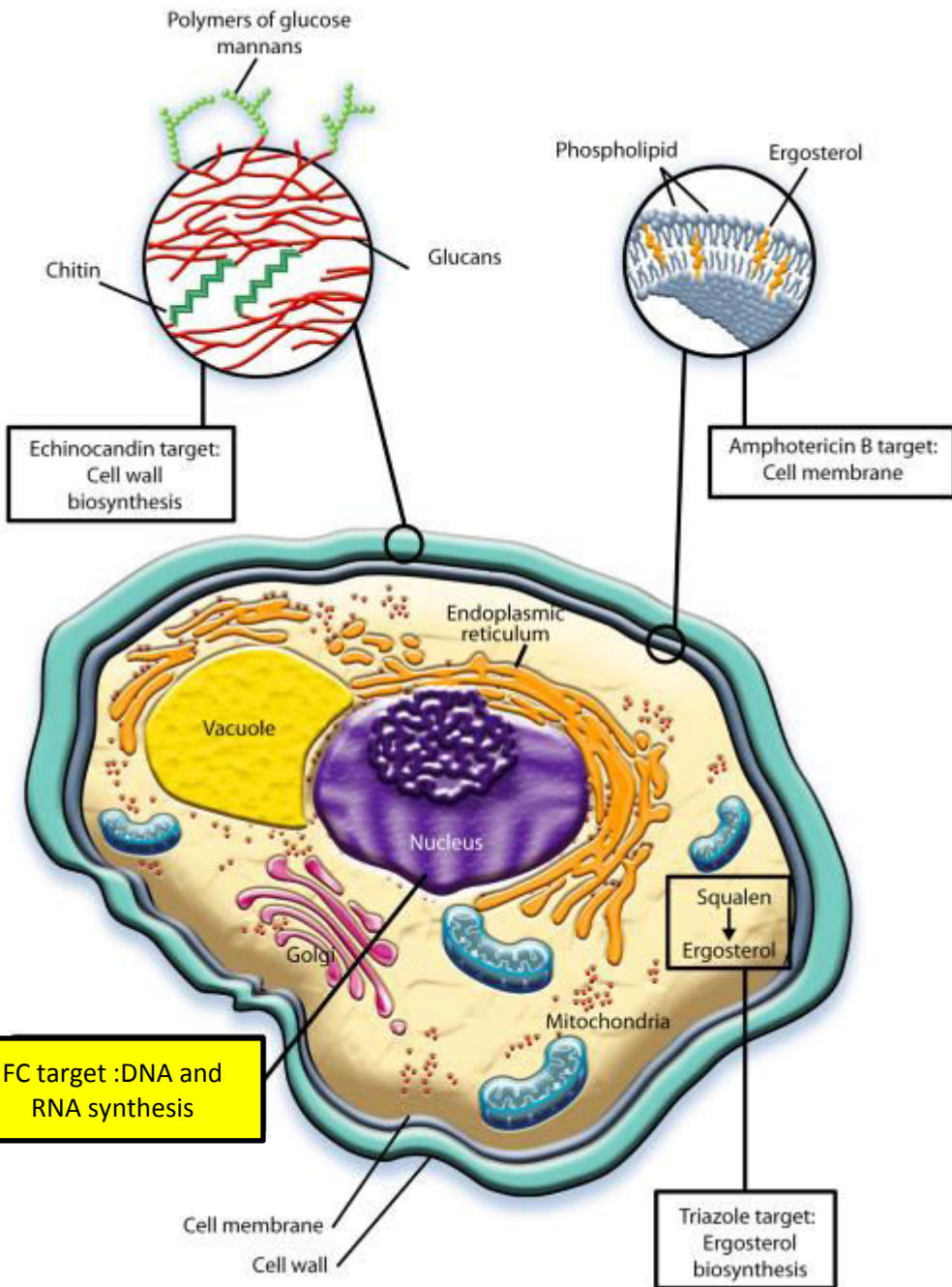
Nikkomycin Z

- **TETRAHYDROFURAN  
DERIVATIVES**

Sordarins, azasordarins

- **OTHER**

Griseofulvin



## Cell membrane

Fungi use principally Ergosterol instead of cholesterol

## DNA Synthesis

Some compounds may be selectively activated by fungi, arresting DNA synthesis.

## Cell Wall

Unlike mammalian cells, fungi have a cell wall

# Resistance terminology

## **Types of antifungal resistance**

- Intrinsic and acquired resistance
- Microbiological and clinical resistance

# Intrinsic resistance and Acquired resistance

- Intrinsic resistance
  - Primary resistance
  - resistance present before exposure to the drug
  - intrinsically resistant and hypersusceptible strains – extremes of a MIC distribution of a random collection of isolates
- Acquired resistance/ secondary resistance
  - Develop. of resistance after exposure to drug
  - Molecular basis of resistance can be explored

# Microbiological resistance and Clinical resistance

- Clinical resistance
  - infection persists despite treatment with an antifungal drug
  - infecting fungus could show normal susceptibility to the agent in vitro
  - antifungal fails to reach the infected site in sufficient quantity
  - Immune system is unable to clear the fungus inhibited in growth by the antifungal

- Microbiological resistance
  - infecting fungus shows *in vitro* resistance
  - patient responds clinically to the treatment
  - Patients own immune system clears the infection
  - agent reaches the target site in unusually high quantity
  - agent interacts synergistically with other molecules at the site of infection

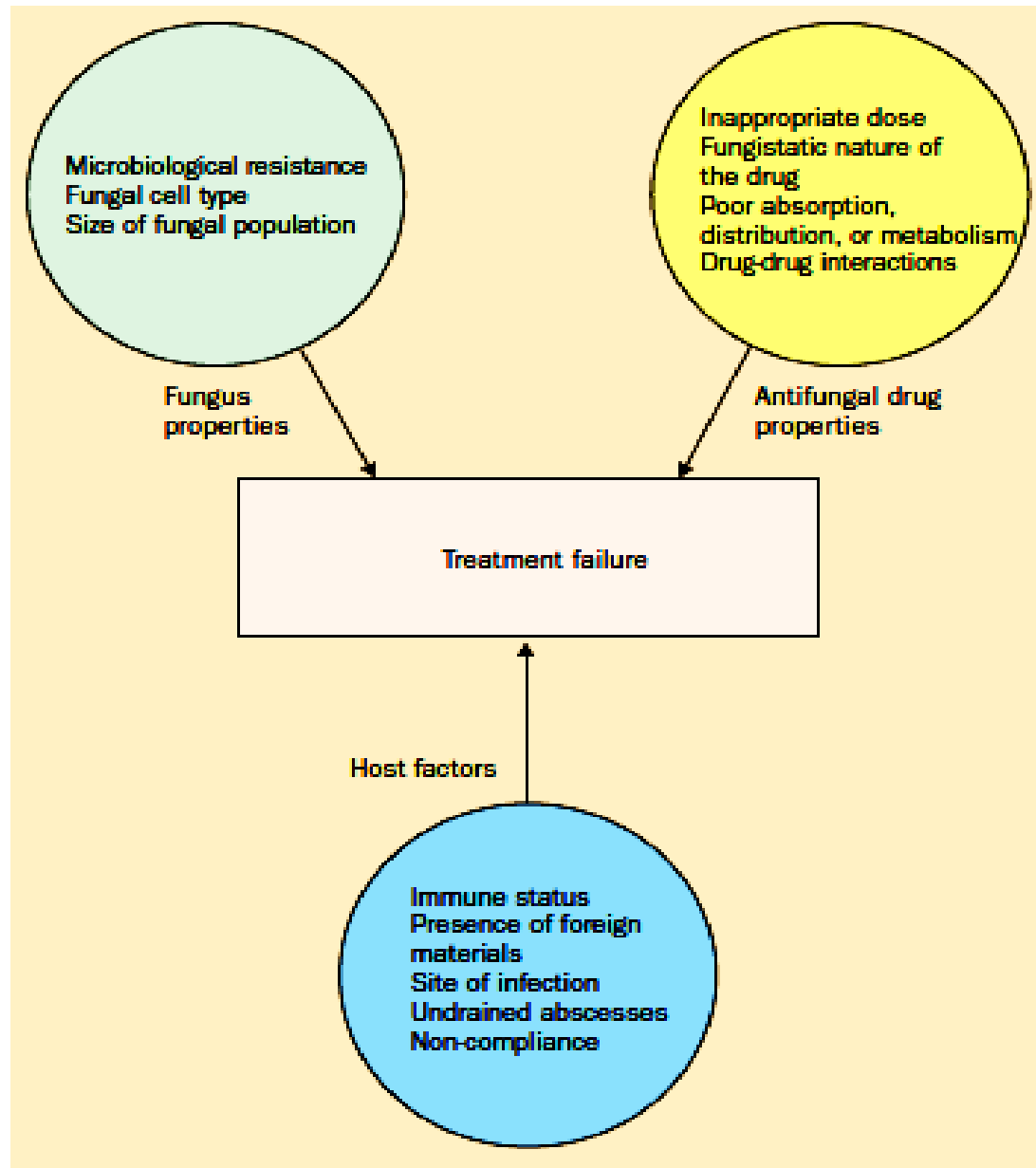
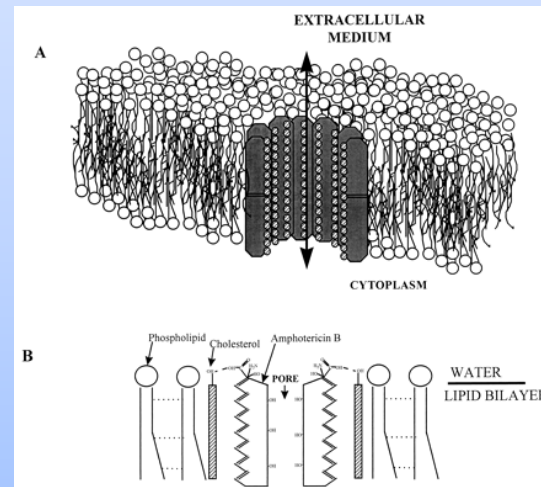


Figure 1. Principal causes of antifungal treatment failure.

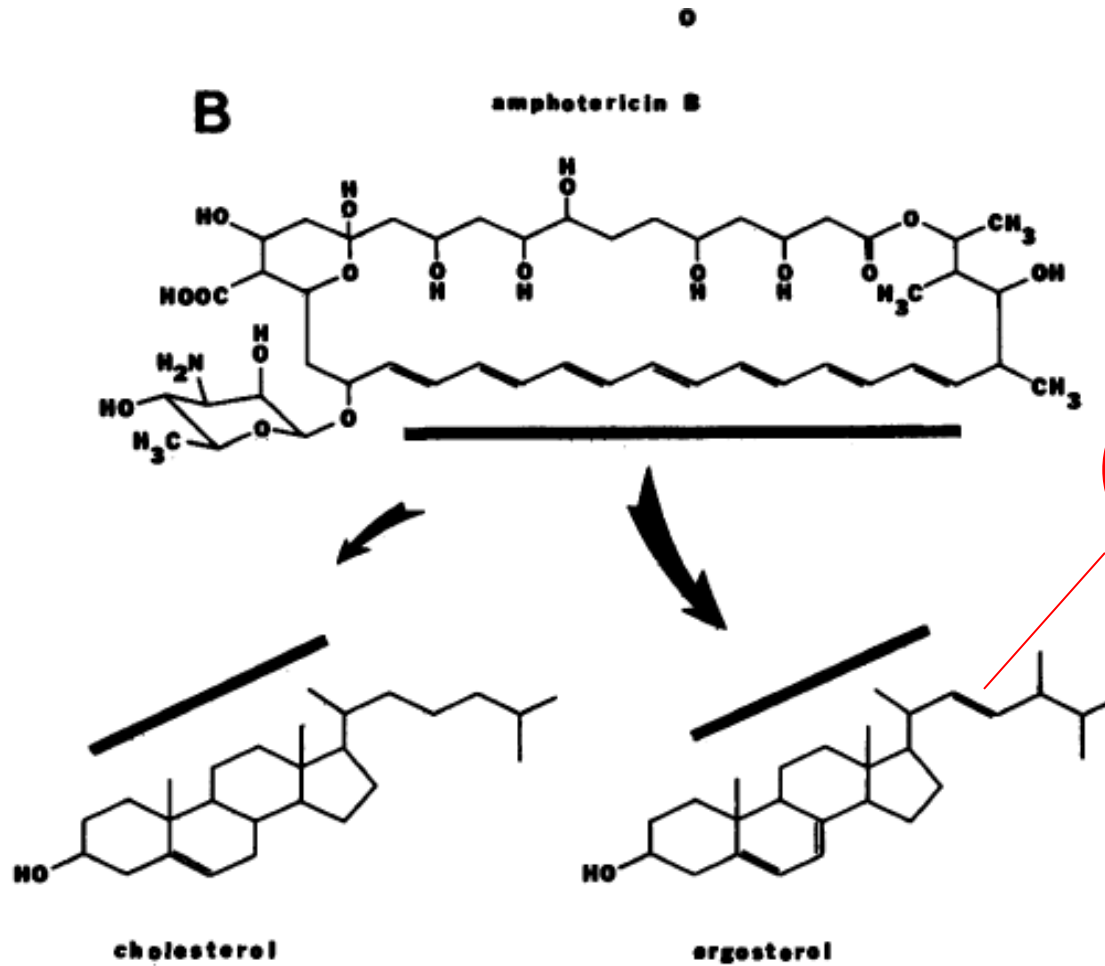


# Amphotericin B

- Binds ergosterol in fungal cell membrane
- Creates transmembrane channel and electrolyte leakage.



# Amphotericin B and ergosterol specificity



# Amphotericin B - Clinical Uses

The drug of choice for:

- Cryptococcal meningitis
- Mucormycosis (zygomycosis)
- Aspergillosis
- Invasive fungal infection, not responding to other therapy

# AmpB resistance

- Intrinsic resistance
  - *Scedosporium spp.*
  - *A. terreus*
  - *Fusarium spp.*
  - *C. lusitaniae*
  - *Trichosporon beigelli*
- Secondary resistance
  - rare
  - Occ. seen in isolates from heavily IC patients

# Mechanisms of amphotericin B resistance

## Amphotericin B resistant fungi

- Quantitative or qualitative alterations in the lipid composition of cell membrane
- ↓ or ↑ ergosterol content limiting the binding of drug

## AMB resistance in *C. lusitaniae*

- Disruption of ergosterol biosynthesis
- *ERG6* a non-essential gene
- Mutation in *ERG6*(Sterol methyltransferase)
  - *erg6* mutant strains
- resistance to Polyenes
- decreased ergosterol content

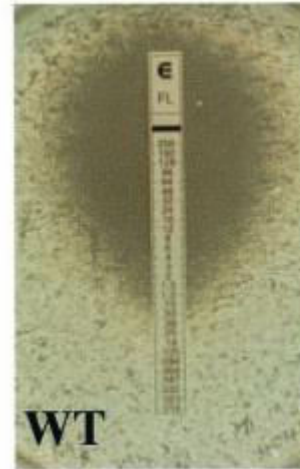


WT



*erg6*Δ

amphotericin B



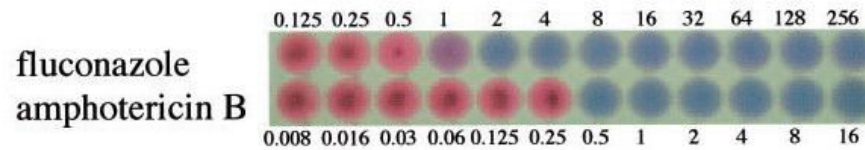
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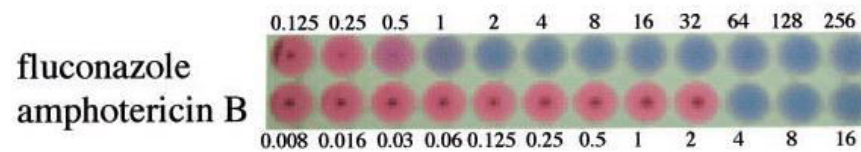
*erg6*Δ

fluconazole

WT



*erg6*Δ



# Flucytosine – Clinical uses

Monotherapy  
- now limited

- Candidiasis
- Cryptococcosis
- Chromoblastomycosis

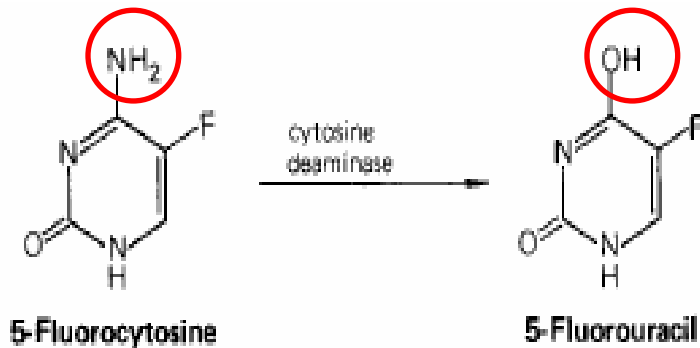
In combination with  
amphotericin B or  
fluconazole.



# Flucytosine

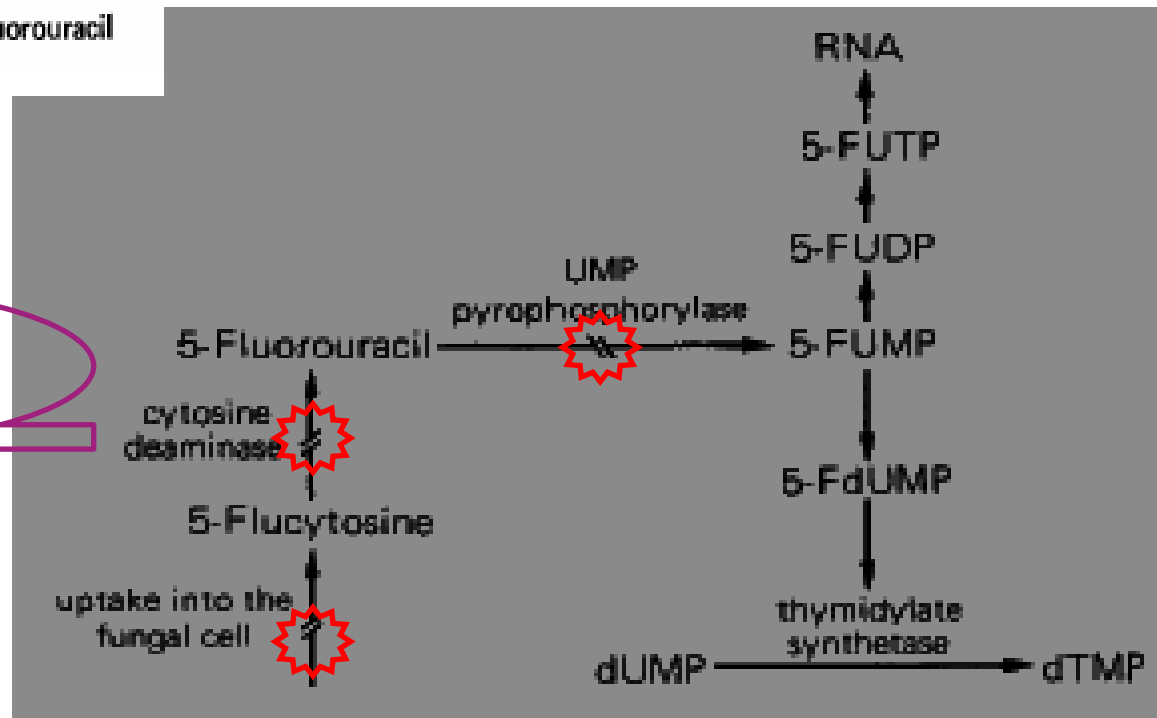
- Acquired Resistance:
  - result of monotherapy
  - rapid onset
  - decreased uptake (loss of cytosine permease activity e.g., *C. glabrata*)
  - altered 5-FC metabolism (loss of cytosine deaminase or UMP pyrophosphorylase activity)  
e.g., *C. albicans* & *Cryptococcus neoformans*

# Flucytosine



Fluorinated pyrimidine related to flurouracil.

Absent in mammals



# Ergosterol Biosynthesis Inhibitors(EBIs)

- Azoles
- Morpholines
- Allylamines
- Thiocarbamates

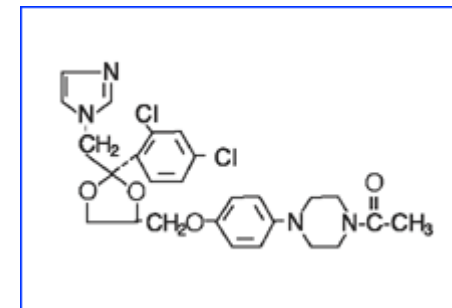
# Azoles

- Most important EBIs
- Demethylation inhibitors (DMIs)

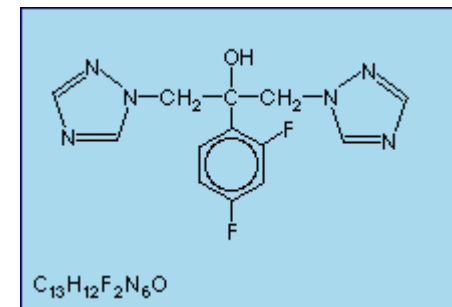
<ul style="list-style-type: none"><li>- Ketoconazole</li><li>- Miconazole</li><li>- Clotrimazole</li></ul>	Imidazoles
<ul style="list-style-type: none"><li>- Fluconazole</li><li>- Itraconazole</li><li>- Voriconazole</li><li>- Posaconazole</li></ul>	Triazoles

## Imidazoles:

- Limited use for treatment of systemic fungal infections
- localized surface infections



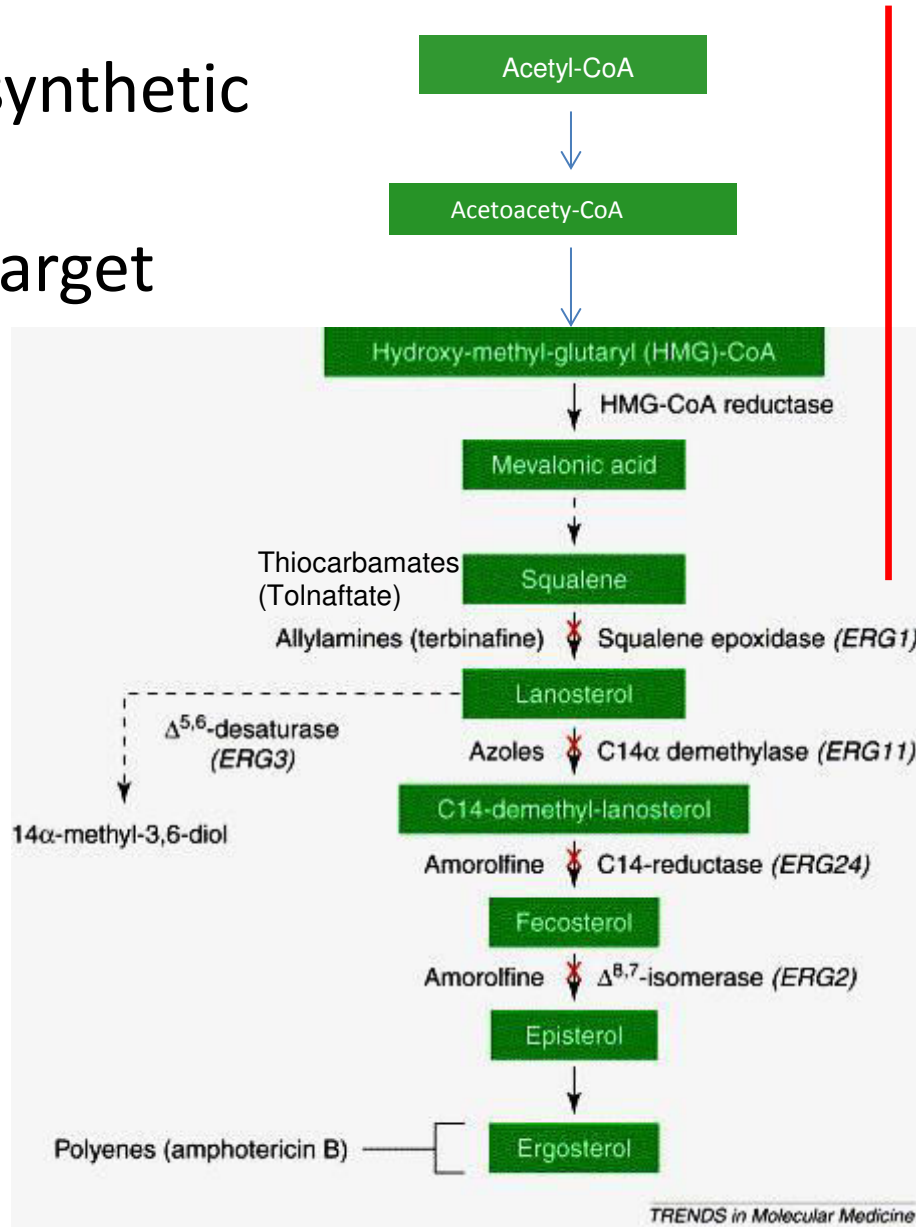
Ketoconazole



C<sub>13</sub>H<sub>12</sub>F<sub>2</sub>N<sub>6</sub>O

Fluconazole

# Ergosterol synthetic Pathway - antifungal target



Early pathway

Late pathway

# Fluconazole - spectrum

- Good activity against *C. albicans* and *Cryptococcus neoformans*
- Non-*albicans Candida* species more likely to exhibit primary resistance

Always resistant



Sometimes resistant

*C. krusei*

>

*C. glabrata*

>

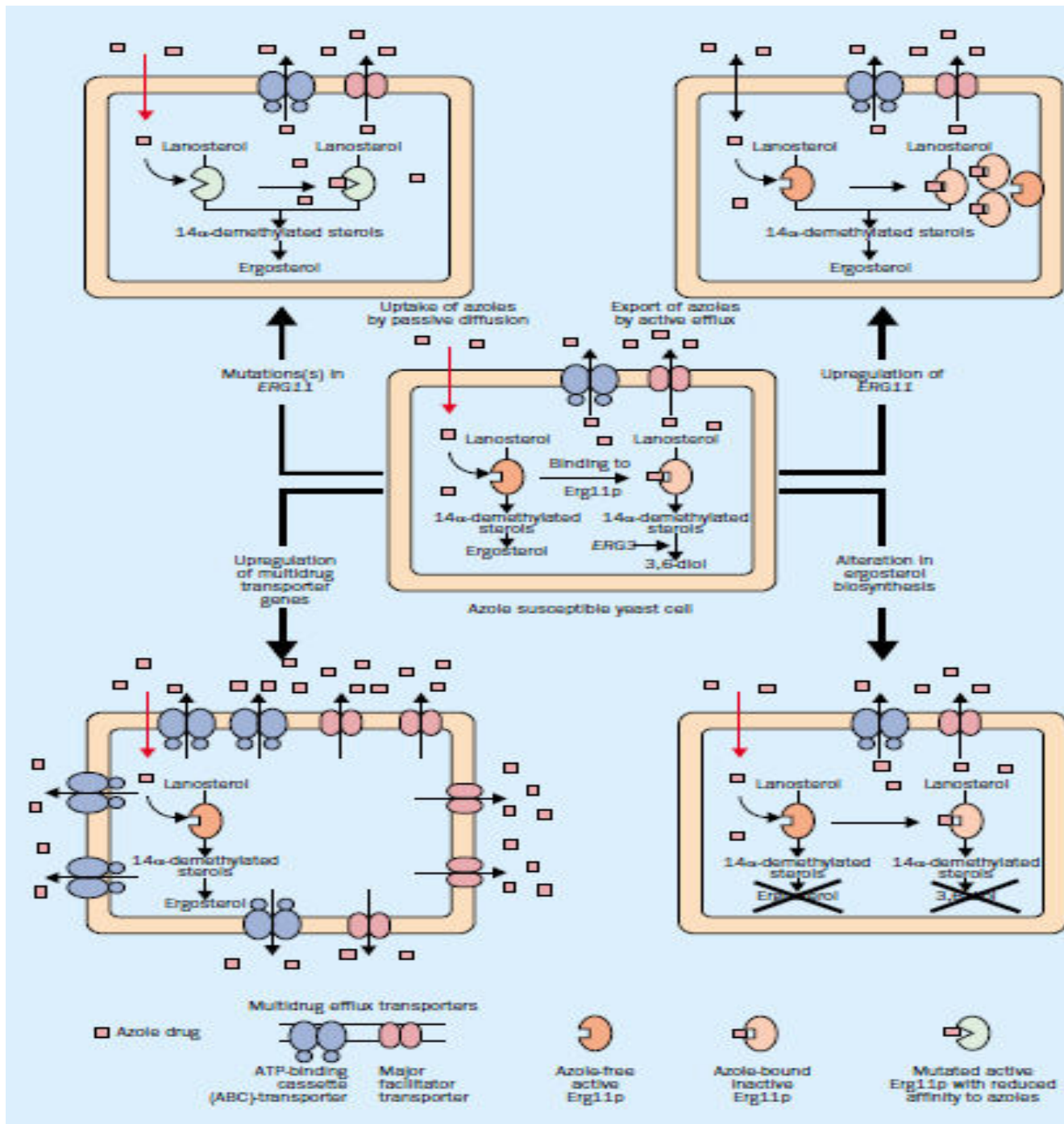
*C. parapsilosis*

*C. tropicalis*

*C. kefyr*

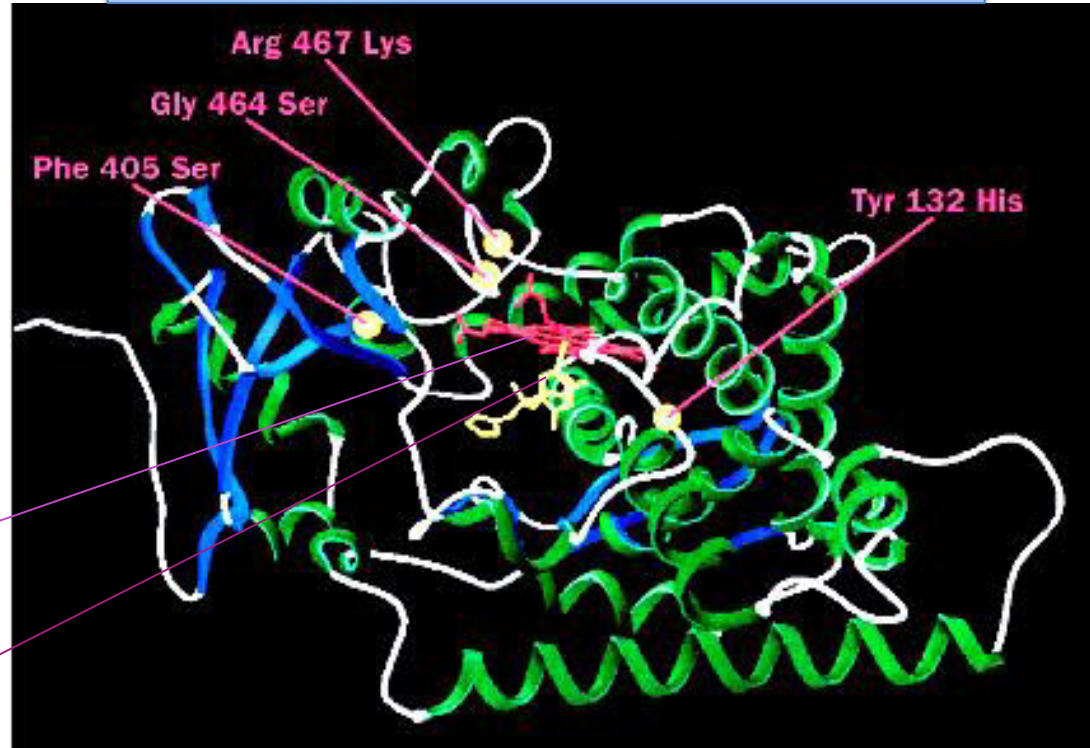
## Mechanisms of azole resistance

- Enhanced efflux of agent
- Alteration of target enzyme
- Overexpression of target enzyme
- Alteration of Ergosterol pathway



Intrinsic resistance to fluconazole in *C. krusei* is due to reduced affinity of Erg11p to the drug as a result of point mutations in it.

Crystal structure of Erg11p (CytP450) depicting Heme - Flu interaction



Heme

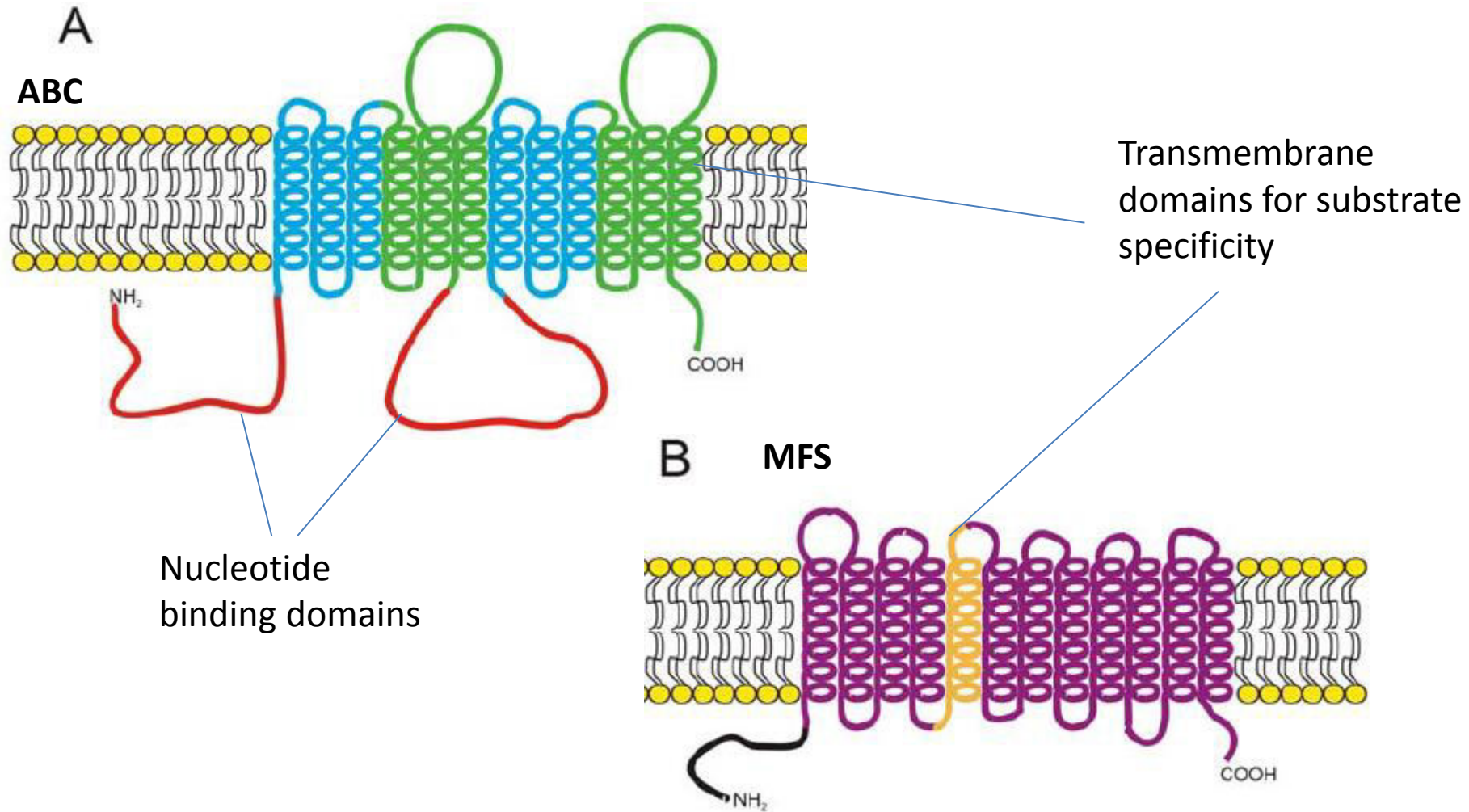
Fluconazole



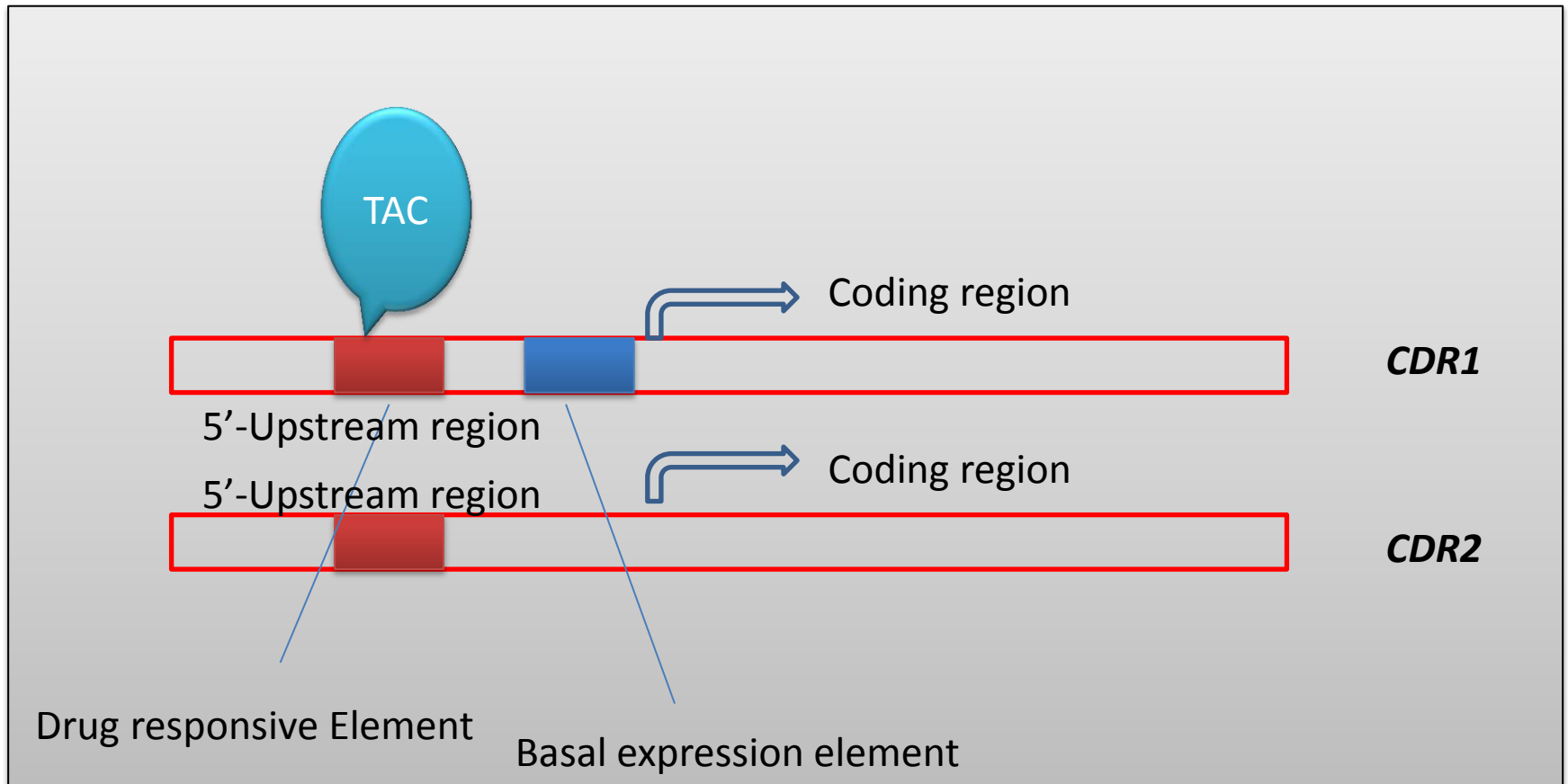
# Drug efflux transporters

- Fungi overcome intracellular toxin accumulation by efflux pumps
  - ABC transporters(ATP driven)
  - MFS transporters(Proton gradient)
- Fungi – 10 to 30 genes encoding transporters per  $10^6$  bp of genome
- MFS transporters – most common in all sequenced fungal genomes

# Structure of ABC and MFS transporters



# Efflux pump transporters in azole resistance in *Candida* spp.



- *CDR1 and CDR2*
  - General phospholipid translocators
  - Fluconazole resistance
  - Most common mechanism of azole resistance in clinical strains of *C. albicans*
- Increased transcriptional activation of *CDR1* and increased mRNA stability in fluconazole resistant isolates of *C. albicans*

Raman *et al*, Antimicrob agents Chemother, 2008

Smirti *et al*, Yeast 2002

Type of resistance	<i>Candida</i> species	Author and date	Name of Journal	principle mechanisms of resistance
Azole resistance	<i>C.tropicalis</i> (n=52)	Jiang et al,2013	J antimicrob Chemother	<ul style="list-style-type: none"> <li>• High <i>ERG</i> 11 expression(n=31)</li> <li>• Miss-sense mutations in <i>ERG</i>11 (12)</li> </ul>
Azole resistance	<i>C.tropicalis</i> (n=1)	Vandeputte et al 2005	Antimicrob agents Chemother	Overexpression of <i>CtERG</i> 11
Azole resistance	<i>Candida</i> spp. (n=4)	Henry et al, 2000	Antimicrob agents Chemother	<ul style="list-style-type: none"> <li>• Global <i>ERG</i> upregulation</li> </ul>

Type of resistance	<i>Candida</i> species	Author and date	Name of Journal	principle mechanisms of resistance
Azole Resistance	C.parapsilosis(n=3)	Silva et al,2011	Antimicrob agents Chemother	Overexpression of MDR1,an MFS transporter
Fluconazole resistance	C.albicans(n=1)	Yan et al,2008	Acta Biochim Biophys sin	Mutation in TAC TF resulting in Cdr1 and Cdr2
Azole resistance	C.albicans(n=2)	Morio et al,2012	J antimicrob Chemother	Sterol $\Delta^{5,6}$ desaturase( <i>ERG3</i> )
Azole resistance	C.albicans	Martel et al,2010	Antimicrob agents Chemother	Mutations in Erg3 (mainly) and Erg11

## Drug resistance in *Cryptococcus neoformans*

- Drug resistance in *Cryptococcus* is a complex problem
- Underlying disease of the patient
- Secondary complications- hydrocephalus, drug intolerance, poor drug compliance, PK issues & development of primary or secondary drug resistance

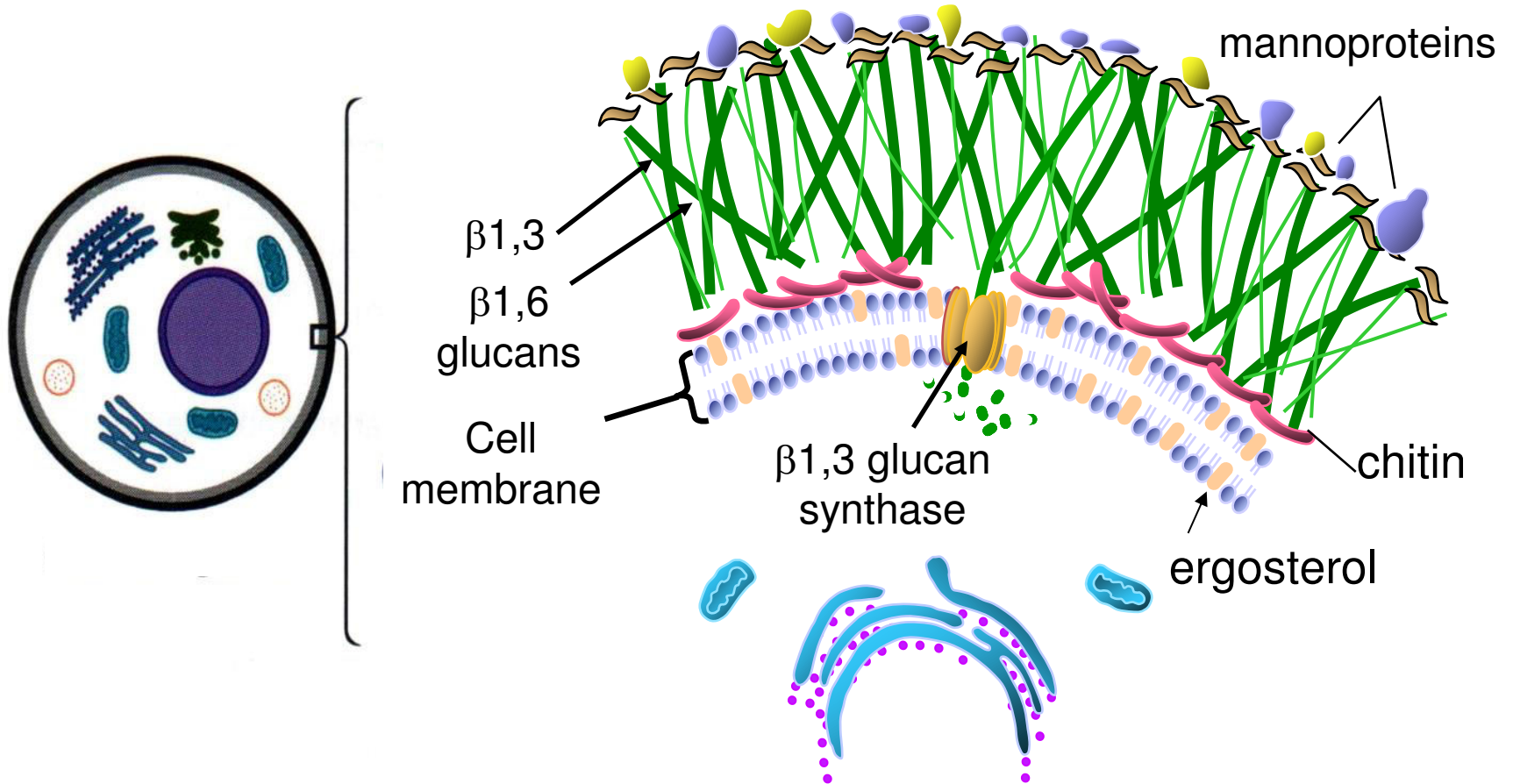
# Drug resistance in *Cryptococcus neoformans*

- Ketoconazole
  - Limited penetration into CNS
  - ineffective in Cryptococcal meningitis
  - successfully used to treat extrameningial cases
- Fluconazole
  - Fails to eliminate infection from genito-urinary tract in disseminated cryptococcosis
  - in vitro resistance due to overexpression of ABC efflux pump Afr1

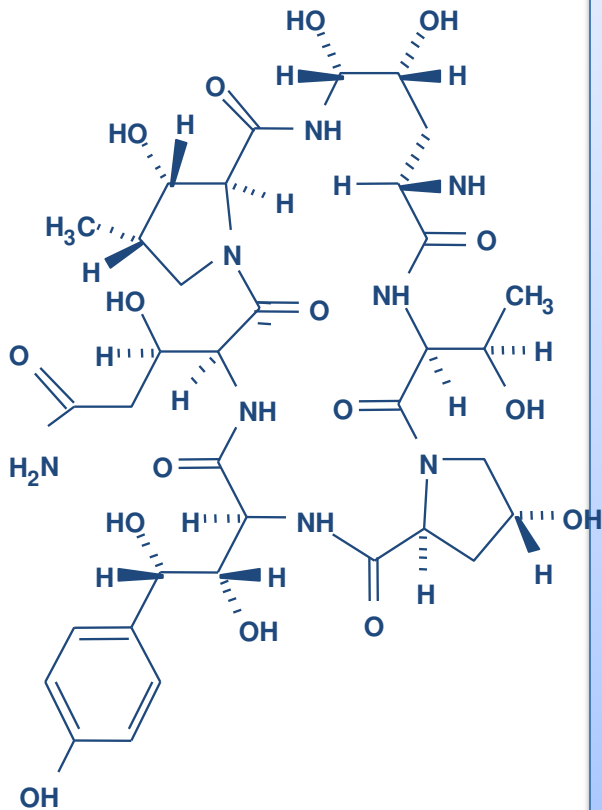


- Amphotericin B failure –
  - direct drug-related nephrotoxicity with the high doses required for treatment of meningitis.
- Innate resistance to echinocandins as *Cryptococcus* has little or no  $\beta(1,3)$ -D-glucan synthase enzyme

# Echinocandins



# Echinocandins - Pharmacology



- Cyclic lipopeptide antibiotics that interfere with fungal cell wall synthesis by inhibition of  $\beta$ -(1,3) D-glucan synthase
- Loss of cell wall glucan results in osmotic fragility

## Spectrum:

- *Candida* species including *non-albicans* isolates resistant to fluconazole
- *Aspergillus spp.* but not activity against other moulds (*Fusarium*, *Zygomycetes*)
- No coverage of *Cryptococcus neoformans*

# Azole resistance in the *A. fumigatus*

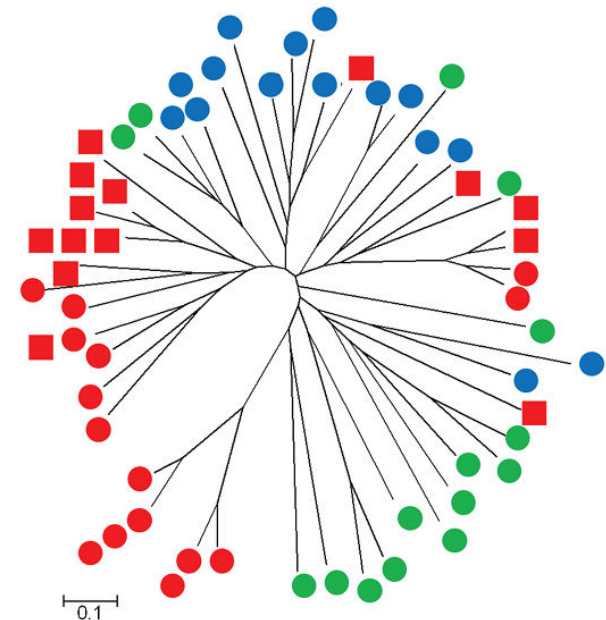
- DMIs have in vitro activity against *A. fumigatus* and their molecule structure are highly similar to clinically licensed triazoles.
- Azole resistant *A. fumigatus* isolates contain an alteration in the target protein sterol 14  $\alpha$  - demethylase (Cyp51) inhibiting drug binding.
- Changes are due to single nucleotide polymorphisms in the gene (*cyp51A*) encoding the protein leading to amino acid substitutions.

# Aspergillosis due to Voriconazole Highly Resistant *Aspergillus fumigatus* and Recovery of Genetically Related Resistant Isolates From Domiciles

Jan W. M. van der Linden,<sup>1,2,a</sup> Simone M. T. Camps,<sup>1,2,a</sup> Greetje A. Kampinga,<sup>3</sup> Jan P. A. Arends,<sup>3</sup> Yvette J. Debets-Ossenkopp,<sup>4</sup> Pieter J. A. Haas,<sup>5</sup> Bart J. A. Rijnders,<sup>6</sup> Ed J. Kuijper,<sup>7</sup> Frank H. van Tiel,<sup>8</sup> János Varga,<sup>9</sup> Anna Karawajczyk,<sup>10</sup> J. Zoll,<sup>1,2</sup> Willem J. G. Melchers,<sup>1,2</sup> and Paul E. Verweij<sup>1,2</sup>

Clinical Infectious Diseases 2013;57(4):513–20

- Study period 1315 *A. fumigatus* isolates from 921 patients screened for resistance.
- Prevalence of azole resistance was 6.8% (63 of 921 patients)
- TR34/L98H - 74.6% (47/63)
- TR46/Y121F/T289A –**20.6%** (13 patients)
- No mutation in Cyp51A - 3 patients



- Microsatellite genotypes of the clinical and environmental resistant TR 46 /Y121F/T289A isolates, compared with TR 34/L98H and wild-type controls

## Azole resistance in the *A. fumigatus*

- Alterations at codon 98 (L98H) of *cyp51A* is predominantly associated with resistance
  - 94% in The Netherlands, 53% in Spain
- Resistance is also associated with an increase in target concentration.
- Strains with L98H also contain a 34-base tandem repeat in the promoter region which causes an eight-fold increase in expression of the *cyp51A* gene (*TR34/L98H*).

## TR34/L98H

- First emerged in 1998 - Dutch patients and endemic in Dutch Hospitals
- Now reported from other parts of Europe, China and India
- Seen both in azole –treated and azole- naïve patients
- Molecular typing studies

**Fungicide driven route of resistance development carries high risk of geographical migration of this resistance trait**

# Mechanism of Resistance Development

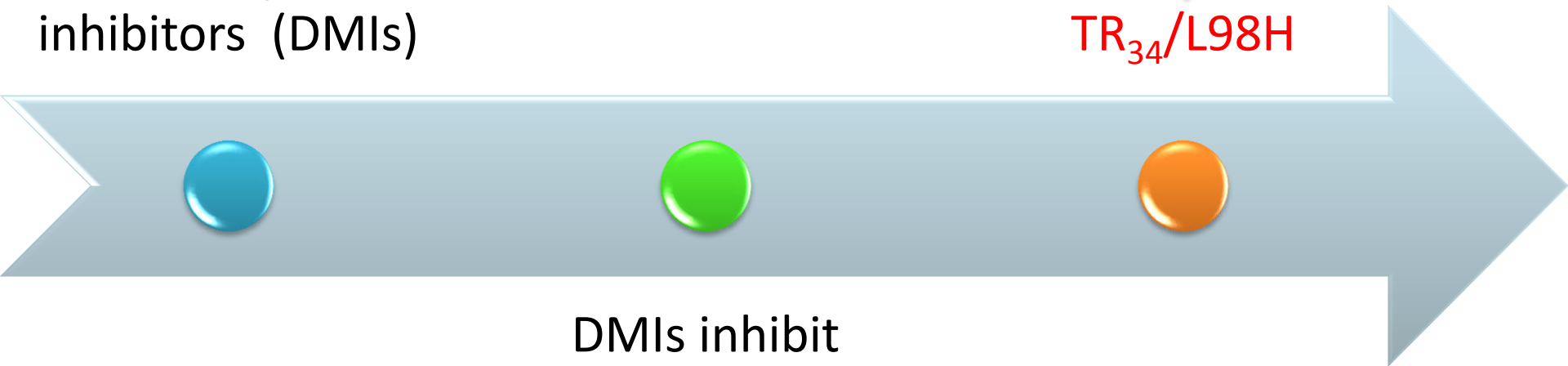
Environmental exposure to 14 $\alpha$  - demethylase inhibitors (DMIs)

Substitution at codon 98 in the *Cyp51A* gene  
+  
34 base-pair tandem repeat in the gene promoter



TR<sub>34</sub>/L98H

DMIs inhibit fungal *Cyp51A* activity



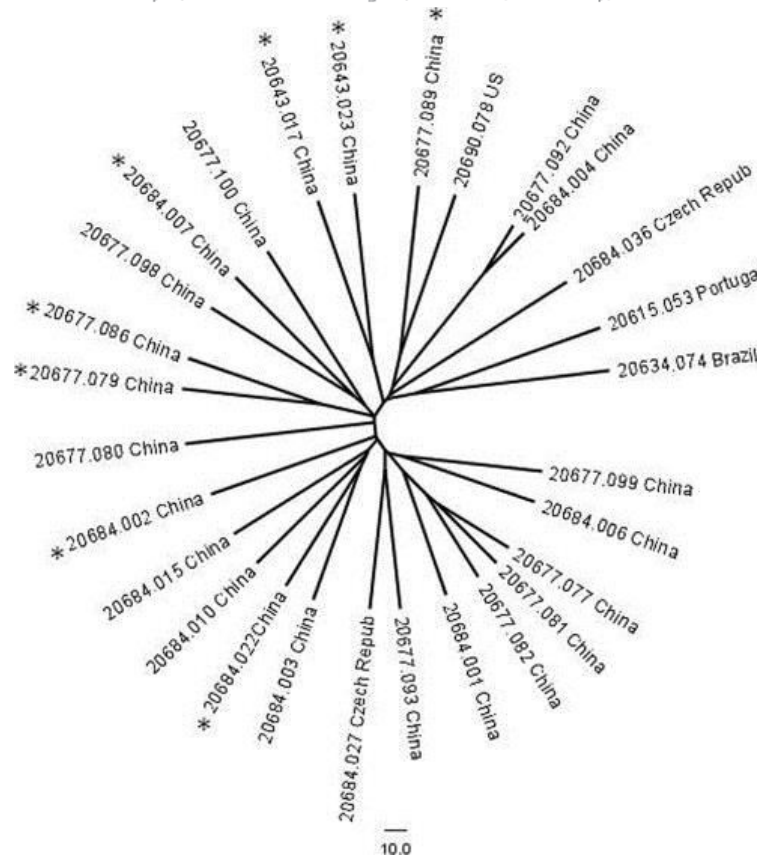


## Azole Resistance in *Aspergillus fumigatus* Isolates from the ARTEMIS Global Surveillance Study Is Primarily Due to the TR/L98H Mutation in the *cyp51A* Gene<sup>▽</sup>

Shawn R. Lockhart,<sup>1\*</sup> João P. Frade,<sup>1</sup> Kizee A. Etienne,<sup>1</sup> Michael A. Pfaller,<sup>2</sup>  
Daniel J. Diekema,<sup>2,3</sup> and S. Arunmozhi Balaje<sup>1</sup>

*Mycotic Diseases Branch, Division of Foodborne, Waterborne and Environmental Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia 30333,<sup>1</sup> and Department of Pathology<sup>2</sup> and Department of Internal Medicine,<sup>3</sup> University of Iowa Carver College of Medicine, Iowa City, Iowa 52242*

28 isolates with high triazole MICs  
From: Brazil, China, Czech Republic, Portugal, USA



Survey of 497 *A. fumigatus*

- Years 2008-2009
- Part of ARTEMIS global surveillance study

Dendrogram of all 28 isolates with elevated triazole MIC values. Isolates with the TR/L98H mutation are marked with an asterisk

# Emergence of New Resistance Mechanism

- In Jan 2010 *A. fumigatus* isolated – Voriconazole well
- MIC of Vori >16mg/L, Itra -2mg/L and posa - 0.5mg/L
- Sequencing analysis of Cyp51A – 2 mutation substitutions  
Y121F and T289A
- 46 bp tandem repeat in promoter gene promoter
- **TR46/Y121F/T289A**

# Echinocandin resistance

- Manipulated or laboratory-selected strains with various degrees of caspofungin resistance have been described.
- They have mutations in the *ECM33 gene* (*AfuEcm33*), encoding cell wall proteins important for fungal cell wall organization.
- Laboratory engineered strains with mutations in the *FKS1 gene* encoding a subunit of the  $\beta$ -1,3-D-glucan synthase enzyme and decreased susceptibility to caspofungin have been generated (Gardiner et al *Med Mycol* 2005)

# Summary of genetic mechanisms leading to antifungal resistance

	Genetic target	
	Candida	Aspergillus
<b>Triazoles</b>		
• Target site mutations	C. albicans	Erg11
• Target site upregulation	C. albicans	Erg11
• Drug efflux transporters		
<b>ABC</b>	C. albicans C. glabrata	Cdr1,Cdr2 Cdr1, Cdr2, Snq2
<b>MFS</b>	C. albicans	Mdr1
• Transcription factors		
<b>ABC</b>	C. albicans C. glabrata	Tac1 Pdr1
<b>MFS</b>	C. albicans	Mrr1
<b>ERG</b>	C. albicans	UpC2
<b>Chromosomal aneuploidy</b>	C. albicans	<b>Chromosome 5</b>
<b>Echinocandins</b>		
Target site mutations	C. albicans C. glabrata	Fks1 Fks1, Fks2